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Nitrous oxide fluxes and nitrogen cycling along a pasture chronosequence in Central Amazonia, Brazil

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Abstract

We studied nitrous oxide (N_2O) fluxes and soil nitrogen (N) cycling following forest conversion to pasture in the central Amazon near Santarém, Pará, Brazil. Two undisturbed forest sites and 27 pasture sites of 0.5 to 60 years were sampled once each during wet and dry seasons. In addition to soil-atmosphere fluxes of N_2O we measured 27 soil chemical, soil microbiological and soil physical variables.

Soil N_2O fluxes were higher in the wet season than in the dry season. Fluxes of N_2O from forest soils always exceeded fluxes from pasture soils and showed no consistent trend with pasture age. At our forest sites, nitrate was the dominant form of inorganic N both during wet and dry season. At our pasture sites nitrate generally dominated the inorganic N pools during the wet season and ammonium dominated during the dry season. Net mineralization and nitrification rates displayed large variations. During the dry season net immobilization of N was observed in some pastures. Compared to forest sites, young pasture sites (≤ 2 years) had low microbial biomass N and protease activities. Protease activity and microbial biomass N peaked in pastures of intermediate age (4 to 8 years) followed by consistently lower values in older pasture (10 to 60 years). The C/N ratio of litter was low at the forest sites (~ 25) and rapidly increased with pasture age reaching values of 60–70 at pastures of 15 years and older.

Nitrous oxide emissions at our sites were controlled by C and N availability and soil aeration. Fluxes of N_2O were negatively correlated to leaf litter C/N ratio, $\text{NH}_4^+\text{-N}$ and the ratio of $\text{NO}_3^-\text{-N}$ to the sum of $\text{NO}_3^-\text{-N} + \text{NH}_4^+\text{-N}$ (indicators of N availability), and methane fluxes and bulk density (indicators of soil aeration status) during the wet season. During the dry season fluxes of N_2O were positively correlated to microbial biomass N, β -glucosidase activity, total inorganic N stocks and $\text{NH}_4^+\text{-N}$. In our study region, pastures of all age emitted less N_2O than old-growth forests, because of a progressive decline in N availability with pasture age combined with strongly anaerobic conditions in some pastures during the wet season.

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1. Introduction

Over the past two decades deforestation rates in the Brazilian Legal Amazon have averaged nearly 20 000 km² per year (Houghton et al., 2000). The majority of the cleared area has been converted into cattle pastures (Nepstad et al., 1997). Clearing of tropical forests for pastures is thought to be an important source of greenhouse gas emissions, such as nitrous oxide (N₂O), methane (CH₄) and carbon dioxide (CO₂) (IPCC, 2001). Studies in tropical America so far have revealed contrasting patterns of N₂O fluxes following forest clearing and pasture establishment. Most studies in Amazonia have shown that (-unfertilized-) pastures emit less N₂O and NO than old growth forest sites (Goreau and de Mello, 1985, 1988; Verchot et al., 1999; Melillo et al., 2001; Garcia-Montiel et al., 2001). Most exceptions to this general finding were measured either on sites which had been fertilized (Luizão et al., 1989; Matson et al., 1990) or were short-term studies measured once during the rainy season (Luizão et al., 1989). The only year-round study where elevated N₂O fluxes were reported from a pasture was the study by Melillo et al. (2001) who measured elevated N₂O fluxes from a newly created pasture in central Rondônia during the first 2 years after establishment. Pastures older than 3 years emitted less N₂O than the forest.

Studies outside Amazonia have shown that compared to original forest, elevated N₂O and NO fluxes may occur in young pastures (Keller et al., 1993; Veldkamp et al., 1999). Pastures older than 10 years all have low to very low emissions (Keller et al., 1993; Veldkamp et al., 1999; Erickson et al., 2001) unless they are fertilized (Veldkamp et al., 1998; Mosier and Delgado, 1997). Davidson et al. (2000) showed that the wide variations in observed N₂O and NO emissions across a range of tropical forest and (-unfertilized-) pasture sites (in Brazil, Costa Rica, and Puerto Rico) were primarily controlled by N availability and soil water conditions. Where N availability was low, N₂O fluxes were also always limited. Where N availability was high, N₂O fluxes were elevated under high soil moisture conditions. How land use change affects N₂O and NO budgets at landscape scale was studied by Plant (2000) and Reiners

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et al. (2002) for NE Costa Rica. Using data-intensive models they showed that the integrated landscape responses depended on both edaphic factors and management.

In the Amazon basin, studies on N₂O emissions together with indices of N cycling have been published so far only from Paragominas, eastern Amazonia (Verchot et al., 1999) and from Rondônia in the south-western Brazilian Amazon (Melillo et al., 2001; Garcia-Montiel et al., 2001). A comprehensive study comparing N₂O fluxes and indices of C and N cycling in pastures of contrasting age has not yet been published for the central Amazon. But we need this additional research to refine our understanding of the magnitude of the changes and the controls on key biogeochemical processes to develop predictions of the consequences of changes in land cover and land use across the Amazon basin. The objective of the present study was to determine the effects of forest-to-pasture conversion and pasture age (time following deforestation) on soil N cycling and related N₂O fluxes near Santarém, Pará in the central Amazon. In our study we used a space-for-time substitution which is a common approach to study the effects of forest clearing and pasture age on trace gas fluxes (e.g. Keller et al., 1993; Veldkamp et al., 1999; Verchot et al., 1999; Melillo et al., 2001; Garcia-Montiel et al., 2001). Space-for-time replications can be an efficient way to generalize qualitative trends and create hypotheses (Pickett, 1989; Fleming, 1999). The main disadvantages of this method involving forest-to-pasture chronosequences have been discussed by Veldkamp et al. (1999): (1) spatial variability is included in the experimental design and may appear as variability caused by time (see also Pickett, 1989); (2) short-term measurements may not be sufficient to derive a good estimate of a parameter; (3) site history is uncertain, i.e. unrecognized effects in the past of a system may be of large magnitude (Pickett, 1989). Because of these problems, Veldkamp et al. (1999) recommended temporal and spatial replication. In the present study we selected a chronosequence of forest (n=2) and pasture sites of contrasting age (n=27). We sampled once during the wet season and once during the dry season at a large number of sites. Our goal was not to provide an accurate estimate of average annual N₂O fluxes from these sites but rather to look for changes in N₂O flux and N availability (and

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their relation to one another). The large number of sites in this chronosequence study would, in part, compensate for the limited temporal sampling.

2. Methods

2.1. Site description

5 The study was conducted near Santarém (2°25' S, 54°47' W) in the state of Pará, Brazil. The landscape consists of a mosaic of old growth forests, pasture land, agricultural fields and secondary forests. The region has a mean annual temperature of 25°C and a mean relative humidity of 86%. Annual rainfall is highly variable and averages about 2000 mm with a dry season lasting usually from August to October (Parrotta et al., 1995). The sites we selected for our study were mainly situated on deep, well-drained clayey Oxisols (kaolinitic yellow latosols in the Brazilian classification system, Embrapa, 1999) with some soils transitional to Ultisols (red-yellow podzolic soils in the Brazilian classification system, Embrapa 1999) also present in the region (Silver et al., 2000).

15 We conducted a regional survey between 50 and 100 km south of Santarém at the beginning of the rainy season in 1999/2000 on a chronosequence of pasture sites (n=27) of contrasting ages (between 6 month to 60 years following deforestation). Two undisturbed old-growth forest sites served as reference plots. Pastures younger than 10 years were established by manual clearing during the dry season. On pastures older than 10 years, bulldozers were also used for removal of trees. Typically, isolated valuable and protected tree species, especially Brazil nut trees, were left in the fields. Slashed and dried biomass was partly burned shortly before the onset of the rainy season. On most sites, grass seeds were either sown directly or after one rotation of maize or rice (10 sites). Typical management of the pasture sites consisted of slash-
20 ing of secondary vegetation once per year and burning about once every three years (n=17) or by a yearly rotation of slashing and burning (n=9). Fire favors the dominance

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of pasture grasses over secondary vegetation and enhances the palatability of grasses. Invading accidental fires from adjacent areas are common. No fertilizers were used on any of the pastures. Except for the 6 month and 1 year old pastures, all sites were actively grazed at an average annual rate of about 1 animal ha⁻¹.

5 *Brachiaria brizantha* (A. Rich.) Stapf cv. Marandu was the dominant grass species at most sites. Only the 1 year old site was planted with “mombaça” (*Panicum maximum* Jacq.) and the 60 year old pasture site was planted with “quicuí” (*Brachiaria humidicola*). All pasture sites of our chronosequence study contained woody vegetation including trees left standing following forest clearing and/or regrowing shrubs. Woody
10 vegetation was more prominent in younger pasture sites where single trees were left standing and residual uncombusted woody debris still remained from the old growth forests.

2.2. N₂O and CH₄ flux measurements

15 Surface fluxes of N₂O and CH₄ were assayed once during both wet and dry season using two-part static vented chambers (PVC and ABS plastics) approximately 20 cm high (Keller and Reiners, 1994). At each site 7–8 chamber bases were installed along at randomly selected points along a 50 m long transect. The chamber bases were inserted into the soil to a depth of 2–3 cm about 5 to 10 min prior to chamber closure. Four 20 ml headspace samples were withdrawn at 10 min intervals over 30 min
20 using nylon syringes fitted with polycarbonate/polypropylene valves. The syringes were transported to the laboratory for analysis by ECD (N₂O) and FID (CH₄) gas chromatography within 36 h of sampling. Integrated peak areas for samples were compared to known standards for determination of gas mixing ratio. Fluxes were calculated from the linear change of mixing ratio with time for the brief enclosures. Fluxes of N₂O were
25 discarded when the initial sample (1 minute following enclosure) greatly exceeded the ambient mixing ratio of ~310 ppbv. When fewer than 4 chamber measurements per site were obtained, the entire site was excluded. Zero fluxes (no change in concentration) were included.

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2.3. Soil sampling and processing

Surface soil (10 cm depth) was sampled once during the rainy season (April–May) and once during the dry season (September–October) of 2000. Soil cores were taken adjacent to 4 chambers with a root corer of 7 cm diameter, transferred in sealed plastic bags and transported in Styrofoam coolers to the laboratory the same day. Coarse roots, stones and litter were manually removed from the samples. Bulk density measurements (0–5 cm depth) were made using undisturbed soil cores. Subsamples for determination of particle size distribution, pH and exchangeable cations were air-dried, ground and passed through a 2 mm sieve. Air-dried subsamples (taking during the rainy season) for total nitrogen (N) and inorganic phosphorus (Pi) were ground to <150 μm using a ball mill. Field moist subsamples (of both wet and dry season sampling) for enzyme analysis (protease activity), microbial biomass N measurement and inorganic N and N-mineralization determination were passed through a 2 mm sieve and stored at 4°C until analysis.

2.4. Soil physical and chemical analyses

Soil bulk density (BD) was measured using the core method (Anderson and Ingram, 1993). Particle size distribution was analyzed with the pipette method using pyrophosphate as a dispersing agent (Moschrefi, 1983). Soil pH was determined in a slurry of 10 g soil and 25 ml de-ionized water (Anderson and Ingram, 1993). Gravimetric water content was calculated as the ratio of mass of water to soil dry mass after oven-drying soil at 105°C for 24 h. Total N was measured with an automated C and N analyzer (Elementaranalysator CNS, Vario EL, elementar, Hanau, Germany) (König and Fortman, 1996). Inorganic P (Pi) was determined by resin extraction (Tiessen and Moir, 1993). Exchangeable calcium (Ca), magnesium (Mg), potassium (K) and aluminum (Al) were extracted by leaching soil samples with 100 ml of unbuffered 1 M ammonium chloride (NH_4Cl) for 4–5 h. Cations in the extracts were quantified by atomic absorption spectroscopy (König and Fortmann, 1996).

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Nitrate (NO_3^-) and ammonium (NH_4^+) were determined by extracting 10 g of field moist soil with 70 ml of 2 M KCl. The soil-KCl solution was shaken for an hour on a horizontal shaker and allowed to settle for 2 h. A 20 ml aliquot of the supernatant was removed with a syringe, filtered through a $0.45\ \mu\text{m}$ polysulfone membrane and stored frozen until analysis (Erickson et al., 2001). Analysis of NH_4^+ and NO_3^- was done on a continuous-flow autoanalyzer (Skalar, Erkelenz, Germany) according to König and Fortmann (1996).

Potential net mineralization and net nitrification were measured using the aerobic incubation procedure described by Hart et al. (1994). Two 10 g sub-samples of field moist soil were transferred into 100 ml polyethylene plastic bottles. One set of sub-samples was immediately extracted with 70 ml of 2 M KCl, the other set was covered with perforated parafilm and incubated for 7 days at ambient temperature in the dark before extraction. Ammonium and NO_3^- were analyzed as described above. Net mineralization and nitrification rates were calculated from the difference between inorganic N at the beginning and end of the incubation, and results were expressed as $\mu\text{g N}$ released per g of dry soil over 7 days.

2.5. Soil microbiological assays

Protease activity (N-benzoyl-L-argininamide hydrolysis) of the soil was determined according to Bonmati et al. (1998). The released NH_4^+ was measured according to the procedure of Kandeler and Gerber (1988). Results of activity are expressed as $\mu\text{g NH}_4^+$ released per g of dry soil per hour.

Beta-glucosidase activity in the soil was measured according to Eivazi and Tabatabai (1988). Toluene was not included in the procedure. Results of enzyme activity is expressed as micrograms p-nitrophenol (PNP) released per g of dry soil per hour and are averages of triplicate assays.

Microbial biomass N was analyzed by the chloroform-fumigation-extraction method of Vance et al. (1987). Soil was adjusted to 30% water holding capacity (WHC) prior to fumigation and extraction. Total organic N of the K_2SO_4 -extracts was oxidized to

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NO_3^- by UV and persulphate digestion, and the released NO_3^- was measured on a continuous flow autoanalyzer (Skalar, Erkelenz, Germany) (König and Fortman, 1996). Microbial biomass N was calculated by subtracting extractable N in the unfumigated sample from extractable N in the fumigated sample (Vance et al., 1987) divided by the K_{EN} factor of 0.54 (Brookes et al., 1985).

2.6. Plant sampling and analysis

Wet and dry season standing grass biomass and grass litter at each pasture site was sampled from 2 points along the transect using a 1 m² frame. Subsamples were dried at 65°C and ground to <150 µm on a ballmill. Total C and N were measured with an automated C and N analyzer (Elementaranalysator CNS, Vario EL, elementar, Hanau, Germany) (König and Fortman, 1996). For comparison to pasture biomass, we used data of forest floor litter stocks including data from Silver et al. (2000).

2.7. Statistical analyses

Fluxes of N₂O and soil variables (log transformed where necessary) were compared using analysis of variance separately for wet and dry seasons with three pasture age classes. Pasture sites were clustered in young (0 to 2 years, n=8), intermediate (4 to 8 years, n=9) and old age classes (>10 years, n=10). The Tukey-Kramer test at the 95% confidence level was used for post-hoc comparison of class means to account for unequal sample sizes.

The relationship between N₂O fluxes and the various soil parameters were evaluated using the linear regression procedures from SYSTAT version 10.2 (SYSTAT, 2002). Number of sites were n=2 for old growth forests, n=3 for 6 month old pastures, n=1 for 1 year old pasture, n=4 for 2 year old pastures, n=2 for 5 year old pastures, n=3 for 6 year old pastures, n=2 for 8 year old pastures, n=4 for 10 year old pasture, n=1 for 12 year old pasture, n=2 for 15 year old pasture, and n=1 for each 20, 25 and 60 year old pastures.

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3. Results

3.1. General soil characteristics of the forest and pasture sites

All sites of our chronosequence study had a clayey to very clayey texture according to the Brazilian classification system (Embrapa, 1999). Clay content varied between 36 and 78% in the top 10 cm of all sites except for one forest site which had a clay content of 25% (Table 1). Soil bulk density (BD) was below 1 Mg m^{-3} at the old growth forest and 6 month old pasture site and increased with pasture age to about 1.2 Mg m^{-3} in the 15 to 25 year old pastures.

Total N of forest and pasture soils up to 8 years old was generally between 2.1 and 2.6 g N kg^{-1} soil (Table 1) and was lower in pastures older than 10 years associated with the higher sand content in older pastures (Table 1).

Soil pH in the forests was 3.9 and between 4.0 and 5.6 in the pasture soils. Forest clearing, burning and pasture establishment increased soil pH and concomitantly decreased available Al at all pasture sites. Concentrations of basic cations Ca (Table 1) and Mg and K (data not shown) increased considerably after deforestation and pasture establishment. Calcium concentrations remained elevated in the pasture sites for up to 12 years after deforestation relative to the forest sites and declined consistently in pastures older than 15 years. Inorganic phosphorous (Pi) was highest in the old growth forest soils and the young pastures of 6 months and 1 year. With increasing pasture age Pi decreased steadily.

3.2. Fluxes of N_2O and CH_4 from forest and pasture soils

Six months following forest clearing and pasture establishment N_2O fluxes were 67% lower compared to the primary forests (Fig. 1). Fluxes of N_2O were higher during the wet than during the dry season and showed no consistent trend with pasture age. During the wet season, pasture sites generally had lower fluxes than forest sites except for the N_2O fluxes of one 6 year old pasture site which exceeded forest N_2O fluxes by

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50%. During the dry season emissions of N_2O from pasture sites were significantly lower than from forest soils and some pasture soils showed slight net uptake of N_2O . Nitrous oxide emissions of the 4 year old pasture sites were comparable to fluxes from the forest sites. Fluxes of N_2O from young, intermediate and old pasture classes were not significantly different from one other.

Forest soils consumed CH_4 during both wet season and dry season at similar rates (Fig. 2). During the wet season young pastures up to one year generally consumed CH_4 whereas most pastures older than 1 year emitted CH_4 to the atmosphere. Emissions of CH_4 were not significantly different among young, intermediate and old pasture age classes. During the dry season forest and most pastures soils consumed CH_4 but no clear trend with pasture age was observed. The uptake rates were similar between pastures and the old growth forest. We found no significant relationships between CH_4 fluxes and indices of N cycling or inorganic N stocks or between CH_4 fluxes and soil moisture, BD or soil chemical characteristics.

3.3. Nitrogen cycling and turnover in forest and pasture soils

Total inorganic N-pools (NO_3^- -N and NH_4^+ -N) were greater during the wet than during the dry season and showed no consistent trend with pasture age (Fig. 3). Nitrate was the dominant form of inorganic N at our old growth forest sites during both wet and dry season. At the pasture sites NO_3^- -N generally dominated the inorganic N pools during the wet season whereas NH_4^+ -N was the dominant form of inorganic N in the pasture soils during the dry season. Extractable NH_4^+ -N did not differ significantly among forest and pasture classes during either wet or dry season. During the wet season, extractable NO_3^- -N concentrations in young pastures of 0–2 years were similar to the forest sites. Pastures older than 4 years had significantly lower extractable NO_3^- -N compared to the forest sites. No significant differences among the pasture classes was found. During the dry season, forest NO_3^- -N concentrations were significantly higher than in pasture soils. No statistically significant differences of NO_3^- -N concentrations

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were found among the pasture classes.

Potential net N-mineralization and net nitrification rates were generally higher during the wet season than during the dry season (Fig. 4). At the forest sites nitrification rates were higher than N-mineralization rates during both wet and dry season. During the dry season, most pastures older than 1 year showed net immobilization and nitrification exceeded N-mineralization. During the wet season no statistically significant differences in potential N mineralization and nitrification rates were found among forest and pasture classes. During the dry season, forest sites had significantly higher N mineralization and nitrification rates than the pasture classes which, in turn, did not differ significantly from one another.

Microbial biomass N during the wet season (Fig. 5) was significantly higher in the 4 to 8 year-old pasture class as compared to the young pasture class (0 to 2 years). However, there were no statistically significant differences among other sampled classes. During the dry season, the microbial biomass N in the forest soils was similar to the intermediate pasture age class of 4 to 8 years whereas both young and old pasture age classes had significantly lower microbial biomass N than the forest sites. No significant differences were found among the pasture age classes.

The protease activity in forest and pasture sites was lower during the wet than during the dry season (Fig. 6). No statistically significant differences were found among forest and pasture classes during either the wet or dry season.

The C/N ratio of litter was low at the forest sites and increased with pasture age up to 10 years (Fig. 7). No clear trend was found in pastures older than 10 years. The C/N ratio increased consistently from about 25 in both the 6 month and 1 year old sites to about 70 in the 15 to 25 year old sites. The forest litter C/N ratio was similar to young pasture sites (0–2 years) whereas pastures older than 4 years had significantly higher C/N ratios than the forest sites. The young pasture age class had significantly lower C/N ratios than the old pasture age class. The increase of the C/N ratio with pasture age was accompanied by a decrease in overall pasture productivity and site degradation as was reflected by sparse soil cover. Total aboveground grass biomass

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decreased from about 10 t ha⁻¹ in the 1 to 10 year old pasture sites to about 5 t ha⁻¹ on average in pastures older than 10 years (data not shown).

4. Discussion

4.1. Changes of N transformation along the forest-to-pasture chronosequence

5 The pasture chronosequence showed a strong decline in most measures of N cycling with time following deforestation. The dominance of nitrate over ammonium pools in our forest soils is consistent with results by Matson et al. (1990) and Neill et al. (1995) for humid tropical forest ecosystems in the central and western Brazilian Amazon. While recognizing that nitrate pool sizes as such do not give information on N transformation
10 rates, nitrate accumulation has been interpreted as an indication of an open soil N cycle (Vitousek et al., 1982). This interpretation was supported by results of our aerobic incubations. In our forest soils, net nitrification exceeded net N-mineralization which indicates the potential for N losses by gaseous emission or leaching (e.g. Vitousek and Sanford, 1986; Matson and Vitousek, 1987). In the pastures of our chronosequence,
15 the internal soil N cycle drastically changed compared to the forest sites. While young pastures still display dominance of nitrate over ammonium in the wet season, the clear drop of nitrate concentrations in the older pastures and the dominance of ammonium in the dry season indicate that the internal N cycle of these pastures is more closed. This is supported by low mineralization and nitrification rates, both of which even became
20 negative (net immobilization, Fig. 4) in a considerable number of pastures.

Increasing N limitation with pasture age was reflected in the decrease in litter quality of *Brachiaria brizantha* with age (Fig. 7), a trend that has also been observed in old pastures in other studies (Castilla and Jackson, 1996; Fernandes et al., 2002). A pattern of increased N limitation with increasing pasture age has also been observed
25 in forest-to-pasture chronosequence studies in Rondônia (Neill et al., 1995, 1999) and in Costa Rica (Veldkamp et al., 1999). Progressive N limitation with pasture age is

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probably caused by a combination of early loss of nitrogen through gaseous emissions and leaching, reduced inputs of nitrogen (e.g. by N fixation) and the immobilization of N by grass litter and roots with a high C-to-N ratio. *Brachiaria brizantha* deposits large quantities of litter and decaying roots of very low N content of 0.5 to 1% (Boddey et al., 1996). Furthermore in pastures, the microbial biomass gradually reworks soil organic matter (SOM) to more recalcitrant forms that cycle N more slowly as compared with SOM in the original forest (Neill et al., 1999).

The strong decline of microbial biomass N following deforestation that we observed has also been reported for young pasture sites near Manaus, in the central Amazon (Luizão et al., 1992). Apart from the direct lethal effects of heat, the effect of burning probably limited microbial growth for some time after felling and burning due to changes in substrate availability (Raison, 1979; Kauffman et al., 1995). Slash burning in Amazonian forests is accompanied by a temporary input of low quality, high C/N debris to soils (Kauffman et al., 1995). The peak of microbial biomass N in the 4 to 6 year old pasture sites (wet season) with a high aboveground biomass production is probably related to the large (-above-) and sub-surface inputs of organic material from shoots, roots, and root residues returning labile C and nutrients to the soil as was found in other studies in Brazil (Boddey et al., 1996; Neill et al., 1999). The aboveground biomass in the intermediate pasture age classes was 12 t ha^{-1} compared to 8 t ha^{-1} in both the young and old pasture sites. The steady decline of the microbial biomass in the pastures of 8 years and older may be a result of pasture degradation caused by the combined effects of reduced nutrient stocks, lower plant biomass and related lower organic matter return with a higher C/N ratio.

Extracellular proteases are involved in the mineralization of organic N compounds to amino acids and subsequent ammonification from amino acids to ammonia and CO_2 (Loll and Bollag, 1983; Kalisz, 1988). In our chronosequence study we observed a close correlation between microbial biomass N and protease activity ($r=0.71$, $p>0.001$, $n=28$). The observed pattern of protease activity was probably also related to changes in substrate quality. As soil proteases are limited by available substrate concentrations

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(Tateno, 1988), high activity in intermediate pastures and decreasing activities in older pasture sites was likely due to changes in availability of nitrogen containing organic compounds as pastures aged. In older pastures with a high litter C/N ratio N is rapidly immobilized by the microbial biomass and, after turnover of the microbial population, may be sequestered into the recalcitrant humic soil pool over time (Hall and Matson, 2003).

4.2. Factors controlling N₂O fluxes from forest and pasture sites

Although we sampled N₂O fluxes only twice, our measured N₂O fluxes correlated with several indices of N cycling, suggesting that N availability exerted an important control on N₂O fluxes. As was found by Erickson et al. (2001, 2002) and Davidson et al. (2000), N₂O fluxes in our study were negatively correlated with the litter quality of the forest and pasture biomass (Fig. 8a). The C/N ratio of litter has been proposed as reasonable proxy for N availability in ecosystems (Davidson et al., 2000) and is an important determining factor whether N is mineralized or immobilized. Litter high in N is associated with fast rates of decomposition and rapid soil N turnover which, in turn, may result in increased emissions of N₂O (Erickson et al., 2001, 2002).

The positive correlation with the ratio of NO₃⁻-N to the sum of NH₄⁺-N and NO₃⁻-N which we observed during the wet season (Fig. 8b) and the positive correlation between fluxes of N₂O and the sum of NH₄⁺-N and NO₃⁻-N during the dry season (Fig. 8c) fit the conceptual model of Firestone and Davidson (1989), Davidson et al. (2000) and Davidson and Verchot (2000). In this model, commonly called “Hole-in-the-Pipe”, the first level of control on N₂O fluxes is the rate of nitrogen cycling through ecosystems. The contribution from nitrification (biological oxidation of NH₄⁺) to N₂O production can be significant at certain sites (Firestone and Davidson, 1989; Hall and Matson, 2003).

Although we found no correlation of N₂O fluxes with WFPS, we think that soil aeration content played an important role in some of the observed N₂O emissions. Negative correlations were found between N₂O fluxes and CH₄ emissions (Fig. 8d) in the wet season. Furthermore, chambers in which we measured the highest methane emis-

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sions often corresponded with low or even negative N₂O emissions (results not shown). These results correspond to findings by Keller et al. (1986) and suggest that at least in some pastures strongly anaerobic conditions prevailed during the wet season, which may have led to reduction of N₂O to N₂. Apart from the reduced N availability in older pastures this may have been an additional cause for the observed low (and sometimes negative) N₂O fluxes.

An additional control on N₂O emissions may have been easily available carbon as was illustrated by the correlation of N₂O emission with β -glucosidase activity in the dry season (Fig. 8f). Beta-glucosidase plays a key role in the decomposition of carbohydrates in soils (Shewale, 1982), and the products of β -glucosidase activity are believed to be an important substrate for microbial metabolism. Especially heterotrophic nitrifying and denitrifying bacteria (which can both produce N₂O) require an organic form of carbon as energy source and may depend on β -glucosidase activity for their carbon supply. The importance of available carbon for N₂O production has been discussed before (Groffman, 1991) and positive correlations between N₂O emissions and β -glucosidase activities have also been established for temperate wetland soils (Freeman et al., 1997; Kang et al., 1998).

4.3. Regional comparison of N₂O-fluxes in the Amazon basin

Forest clearing can result in a pulse of available soil N and may cause transient increases in N₂O emissions (Keller et al., 1993; Melillo et al., 2001). The magnitude and duration of N₂O emissions following deforestation and pasture establishment differs among the central and eastern Brazilian Amazon on one hand and the western Brazilian Amazon on the other hand. Our results from 2 forest sites and 27 pasture sites in the central Amazon agree with other studies conducted in the central and eastern Amazon (Goreau and de Mello, 1985, 1988; Verchot et al., 1999); N₂O fluxes from forest soils were relatively high and N₂O fluxes from pasture soils were always lower. The dominance of nitrate over ammonium and the dominance of nitrification over N-mineralization during the wet season (Figs. 2 and 3) may indicate that in pastures

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younger than 10 years there is some potential for higher N₂O emissions. However, we only measured high N₂O fluxes on one 6 year old pasture site.

Our results agree also with observations from Rondônia, western Amazon for pastures older than 3 years (Garcia-Montiel et al., 2001) but do not agree with higher emissions from young pastures up to 2–3 years relative to forest (Melillo et al., 2001). The temporary increases of N₂O in Rondônia immediately following forest clearing may be explained by different soil conditions. In contrast to the clay-textured Oxisols in central and eastern Amazon, soils in Rondônia are coarser textured Ultisols. This difference in texture may lead to rapid SOM turnover and N-mineralization rates in Rondônia (Desjardins et al., 1994; Moraes et al., 1996; Neill et al., 1999), whereas in the eastern Amazon, the higher clay content may protect SOM against rapid mineralization (Koutika et al., 1997). Furthermore, the variation in soil mineralogy on P availability likely contributed to the differences in pasture productivity and N cycling across the Amazon basin (see Davidson et al., 2004). The phosphorus status of pastures on Ultisols in Rondônia is better (see Garcia-Montiel et al., 2000) than the pastures in this study (Table 1) or a nearby study on forest-to pasture chronosequences (Asner et al., 2004), and in the Paragominas region, eastern Amazon (see Verchot et al., 1999). The better P status at the Rondônian sites may have led to higher pasture productivity and hence, the potential of increased levels of SOM and N-mineralization. In contrast to Rondônia, studies in Paragominas (Trumbore et al., 1995) and central Amazonia (Wick, unpublished results) show declining SOM levels in (-unfertilized-) pastures.

Overall, this study confirms the results from a forest-to-pasture chronosequence in Paragominas, eastern Amazon on similar soils and management regimes (Verchot et al., 1999) that young pastures do not emit higher N₂O compared to the forests that they replaced. Excess N, easily available carbon and soil water content controlled fluxes of N₂O.

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was done as part of the Brazilian led Large Scale Biosphere-Atmosphere Experiment in Amazonia (LBA). We also thank E. Davidson for the constructive review of an earlier version of this manuscript.

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Table 1. Average (\pm standard error) soil physical and chemical characteristics of the top 10 cm of old growth forests and pasture soils during the wet and dry season of 2000, near Santarém, Pará, Brazil.

	Clay %	Sand %	BD Mg m ⁻³	WFPS %		pH H ₂ O	Ntot g kg ⁻¹	Pi μg g ⁻¹	Al cmol _c kg ⁻¹	Ca
				wet	dry					
Forest	78	8	0.76	55	43	3.9	2.6	12.4	4.4	0.2
Forest	25	58	ND	ND	46	3.9	1.6	8.6	1.2	0.9
0.5 yr	67	20	0.88	47	42	5.5	2.4	18.1	1.0	5.2
0.5 yr	53	32	0.97	59	40	4.9	2.1	23.0	1.3	3.5
0.5 yr	69	7	0.75	41	35	4.3	2.4	7.8	3.1	1.5
1 yr	66	24	1.06	67	ND	4.0	2.4	13.1	2.9	0.5
2 yrs	72	14	1.11	93	66	4.5	2.4	10.0	3.9	1.9
2 yrs	73	9	1.03	73	55	5.5	2.6	6.1	0.4	6.4
2 yrs	57	24	0.91	40	35	5.2	2.6	7.1	0.8	5.6
2 yrs	72	12	1.05	76	52	5.1	2.4	7.8	0.9	3.2
4 yrs	64	12	0.92	70	53	5.7	2.1	5.8	0.5	7.1
4 yrs	61	11	1.11	83	83	5.1	2.3	6.2	0.3	4.1
5 yrs	62	6	0.98	76	59	5.2	2.3	5.7	0.6	4.6
5 yrs	58	15	0.99	57	48	5.1	2.3	6.9	0.7	4.1
6 yrs	66	18	1.02	72	50	5.1	2.1	9.7	1.4	4.9
6 yrs	57	18	0.91	58	41	5.3	2.5	9.7	0.2	6.3
6 yrs	55	16	1.13	84	75	5.2	2.1	8.3	0.6	3.5
8 yrs	76	7	0.87	62	46	5.0	2.6	11.7	1.0	5.3
8 yrs	47	27	0.98	63	55	4.6	2.1	8.6	1.3	3.0
10 yrs	36	55	1.07	52	28	4.9	1.6	5.7	0.9	2.3
10 yrs	45	45	1.48	92	57	4.9	1.5	6.0	0.9	1.4
10 yrs	72	8	0.93	62	46	4.8	2.0	9.2	0.8	2.5
10 yrs	54	30	1.08	74	47	4.9	2.3	11.8	1.0	4.2
12 yrs	46	25	1.10	66	48	5.5	2.1	6.7	0.1	5.2
15 yrs	43	44	1.20	100	53	4.9	1.8	6.9	0.6	2.0
15 yrs	43	27	1.21	70	55	5.6	1.7		0.1	5.6
20 yrs	40	49	1.17	46	36	5.2	1.6	5.7	0.3	2.1
25 yrs	53	31	1.22	74	78	4.9	1.6	6.5	0.4	1.7
60 yrs	45	45	1.44	91	49	4.8	1.3	5.0	1.0	0.7

ND=not determined.

BGD

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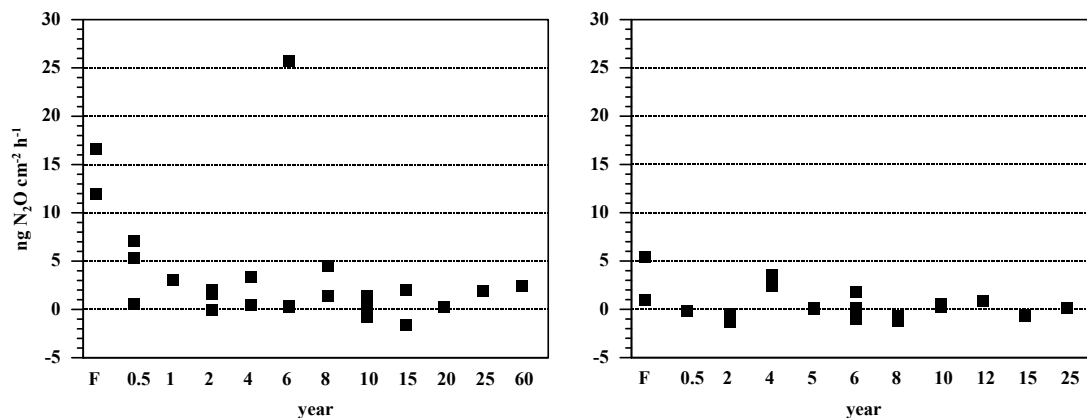


Fig. 1. Fluxes of N₂O (ng N₂O-N cm⁻² h⁻¹) from forest and pasture soils during the wet season (left) and the dry season (right) of 2000, near Santarém, Pará, Brazil.

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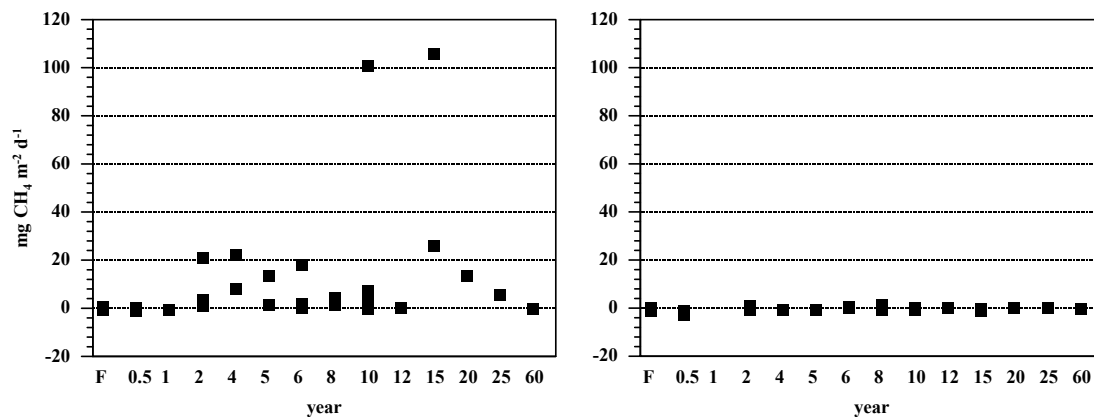


Fig. 2. Fluxes of CH₄ (mg m⁻² d⁻¹) from forest and pasture soils during the wet season (left) and the dry season (right) of 2000, near Santarém, Pará, Brazil.

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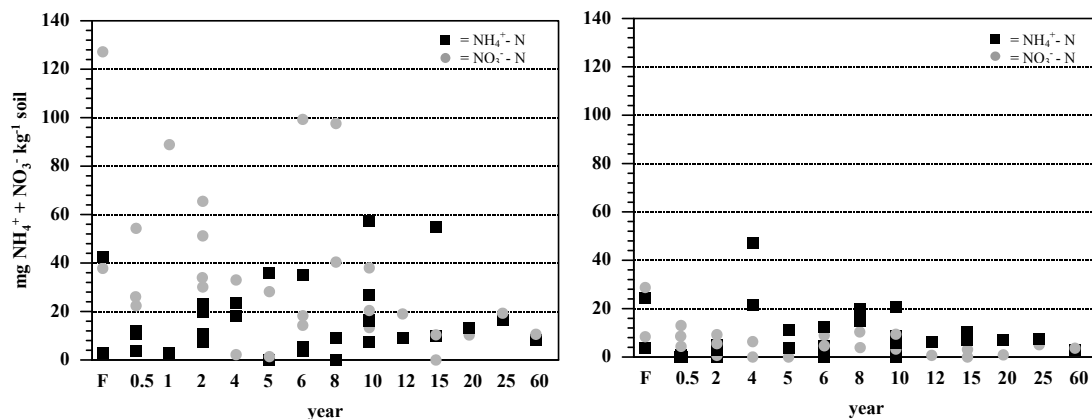


Fig. 3. Inorganic N-pools (mg NH₄⁺ and NO₃⁻-N per kilogram soil) from forest and pasture soils during the wet season (left) and the dry season (right) of 2000, near Santarém, Pará, Brazil.

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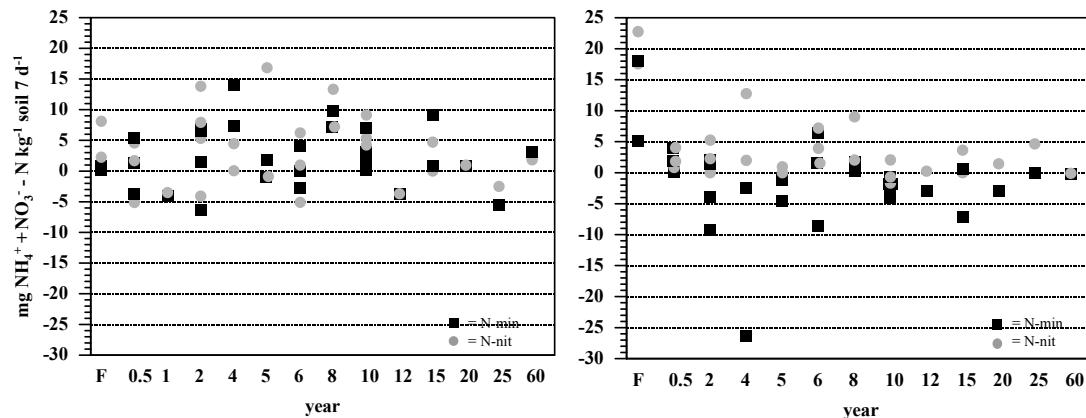


Fig. 4. Potential N-mineralization (mg NH₄⁺ and NO₃⁻-N) and nitrification rates (mg NO₃⁻-N) over 7 days in the laboratory from forest and pasture soils during the wet season (left) and the dry season (right) of 2000, near Santarém, Pará, Brazil.

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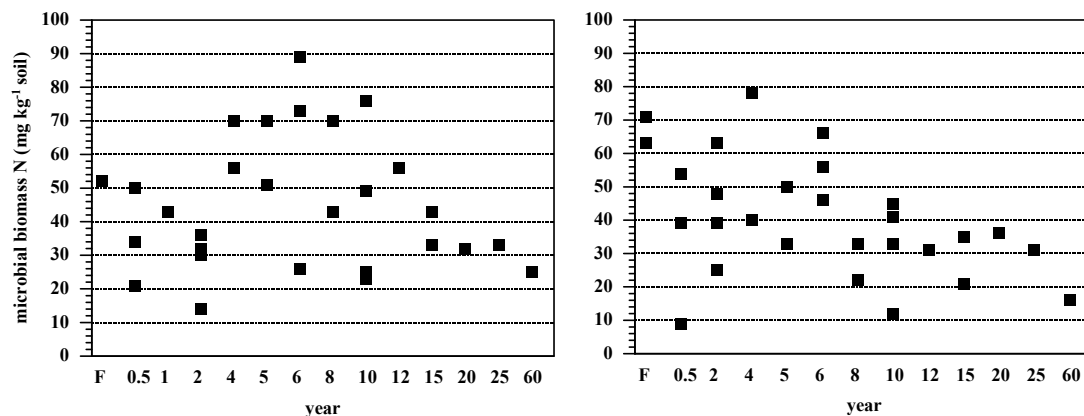


Fig. 5. Microbial biomass N (mg N kg⁻¹ soil) from forest and pasture soils during the wet season (left) and the dry season (right) of 2000, near Santarém, Pará, Brazil.

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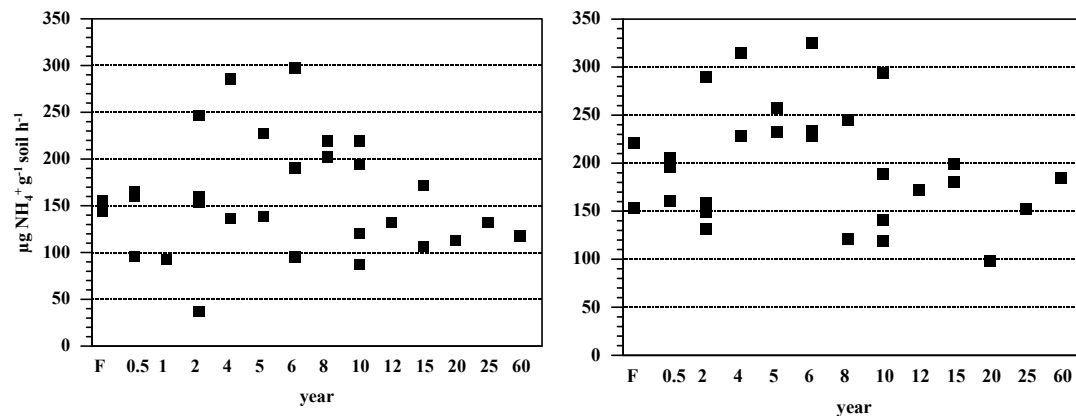


Fig. 6. Protease activity ($\mu\text{g NH}_4^+\text{-N g}^{-1}\text{ soil h}^{-1}$) from forest and pasture soils during the wet season (left) and the dry season (right) of 2000, near Santarém, Pará, Brazil.

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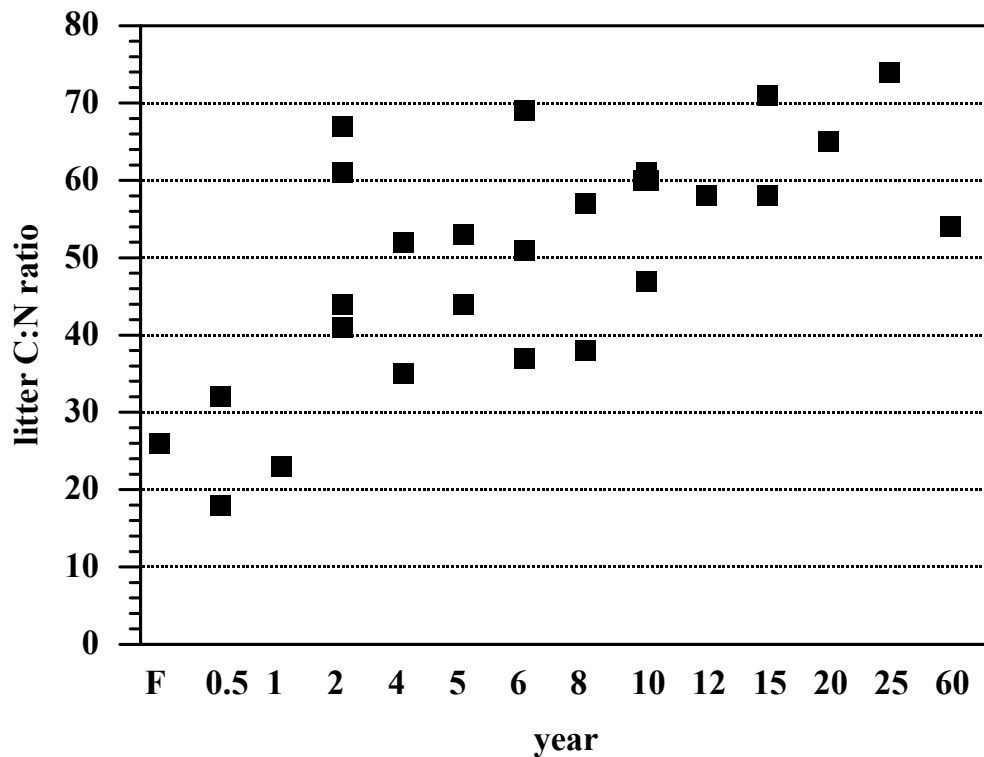


Fig. 7. Litter C/N ratio from forest and pasture soils of 2000, near Santarém, Pará, Brazil.

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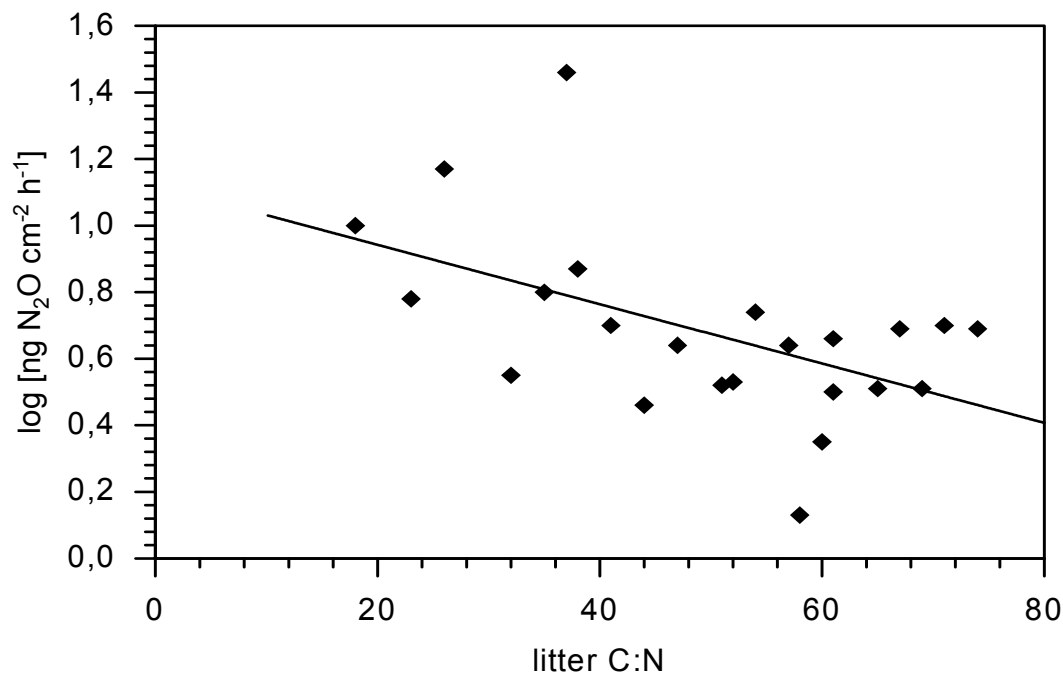


Fig. 8. (a) Linear regressions of wet season log transformed N₂O fluxes with litter C/N ratio [$\log \text{N}_2\text{O} = -0.009 \cdot \text{litter C:N} + 1.12$, $R^2 = 0.28$, $p = 0.01$, $n = 23$] during 2000 from a forest-to-pasture chronosequence near Santarém, Pará, Brazil.

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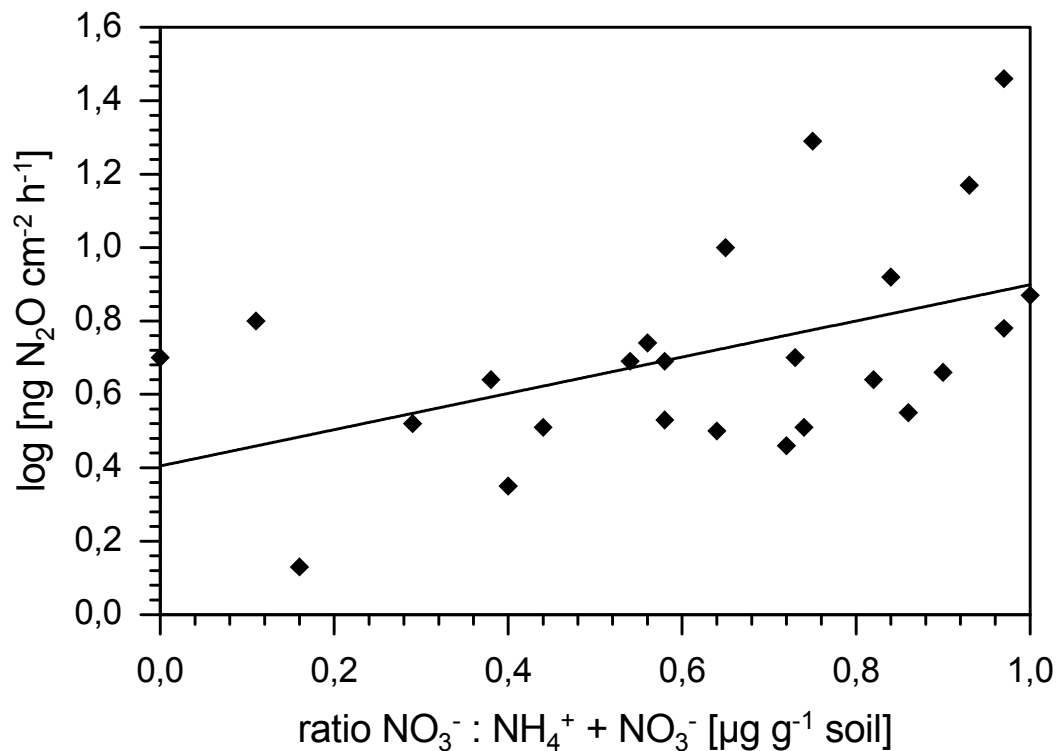


Fig. 8. (b) Linear regressions of wet season log transformed N_2O fluxes with the ratio $\text{NO}_3^- : \text{NH}_4^+ + \text{NO}_3^-$ [$\log \text{N}_2\text{O} = 0.497 \cdot \text{ratio NO}_3^- : \text{NH}_4^+ + \text{NO}_3^- + 0.403$, $R^2 = 0.23$, $p = 0.016$, $n = 25$] during 2000 from a forest-to-pasture chronosequence near Santarém, Pará, Brazil.

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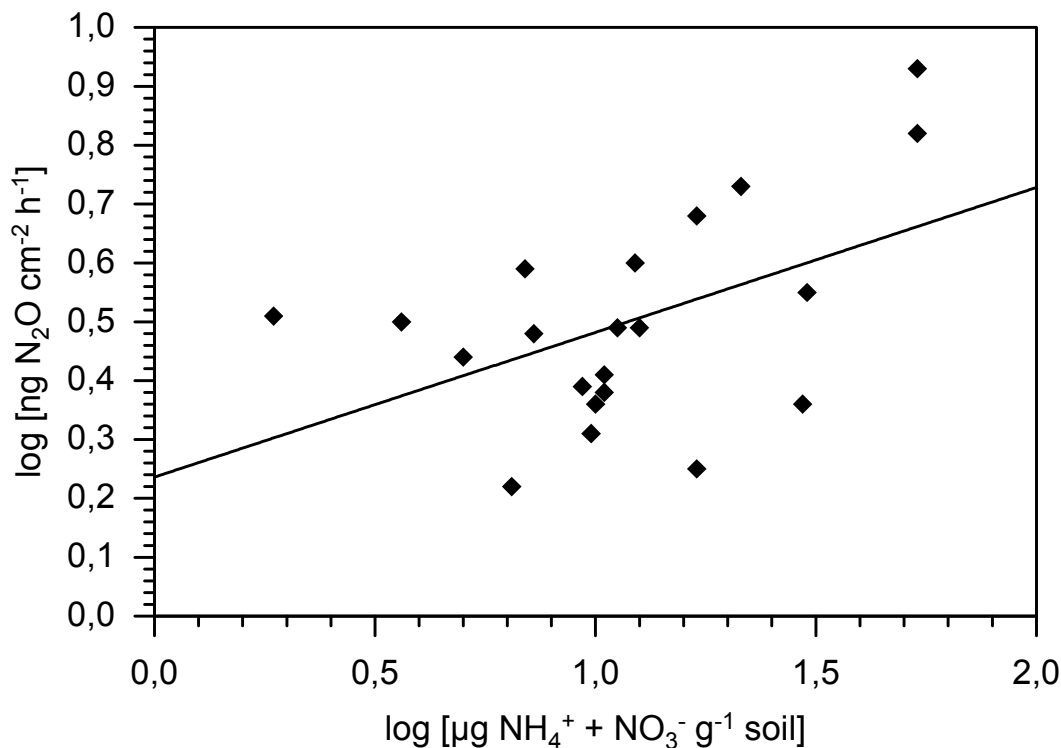


Fig. 8. (c) Linear regressions of dry season log transformed N_2O fluxes with $\log \text{NH}_4^+ + \text{NO}_3^-$ [$\log \text{N}_2\text{O} = 0.244 \cdot \log (\text{NO}_3^- + \text{NH}_4^+) + 0.239$, $R^2 = 0.23$, $p = 0.026$, $n = 21$] during 2000 from a forest-to-pasture chronosequence near Santarém, Pará, Brazil.

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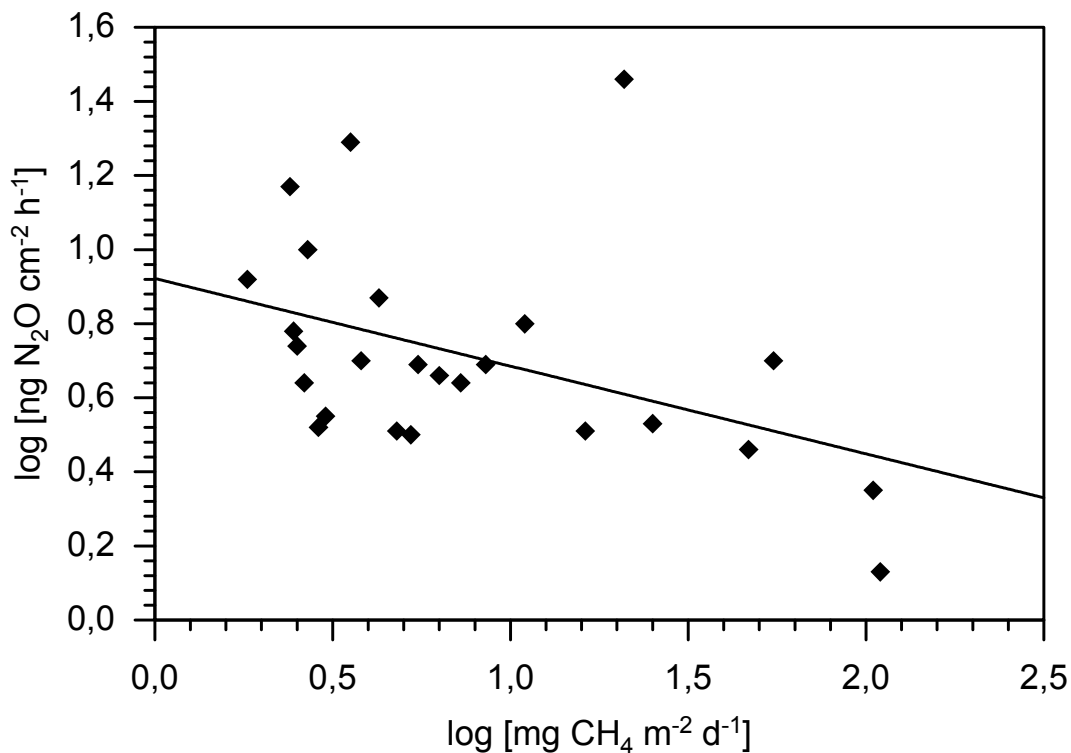


Fig. 8. (d) Linear regressions of wet season log transformed N_2O fluxes with $\log \text{CH}_4 + 50$ [$\log \text{N}_2\text{O} = -0.239 \cdot \log \text{CH}_4 + 0.92$, $R^2 = 0.19$, $p = 0.03$, $n = 25$] during 2000 from a forest-to-pasture chronosequence near Santarém, Pará, Brazil.

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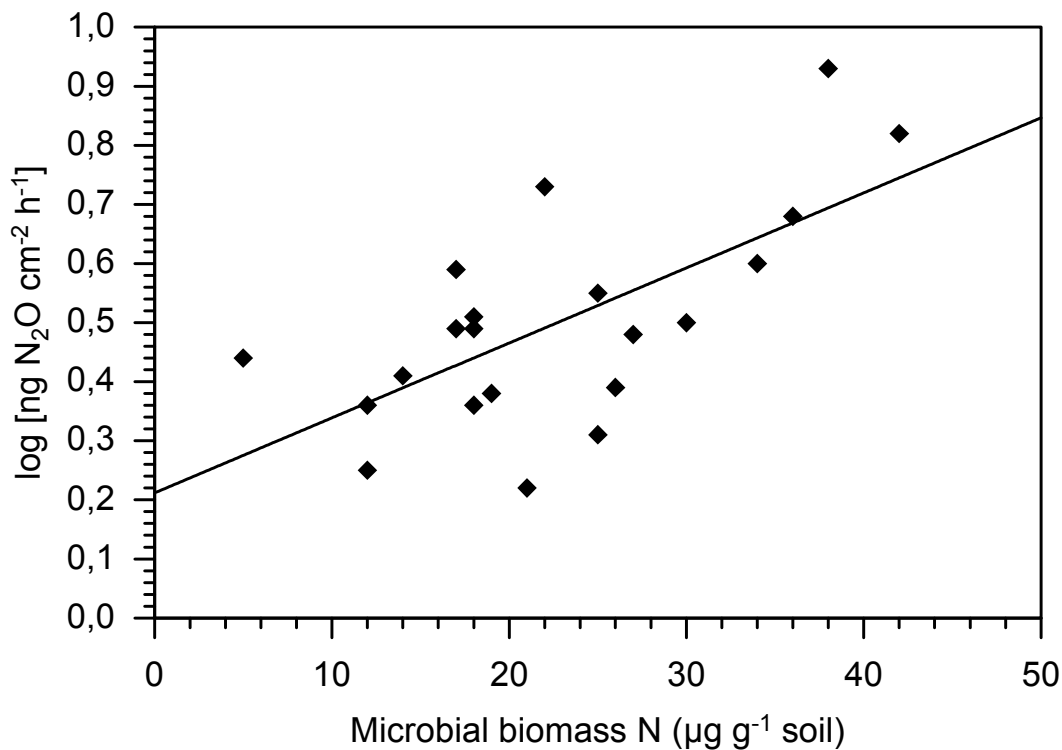


Fig. 8. (e) Linear regressions of dry season log transformed N₂O fluxes with microbial biomass N [$\log \text{N}_2\text{O} = 0.012 \cdot \text{N}_{\text{mic}} + 0.221$, $R^2 = 0.43$, $p = 0.001$, $n = 21$] during 2000 from a forest-to-pasture chronosequence near Santarém, Pará, Brazil.

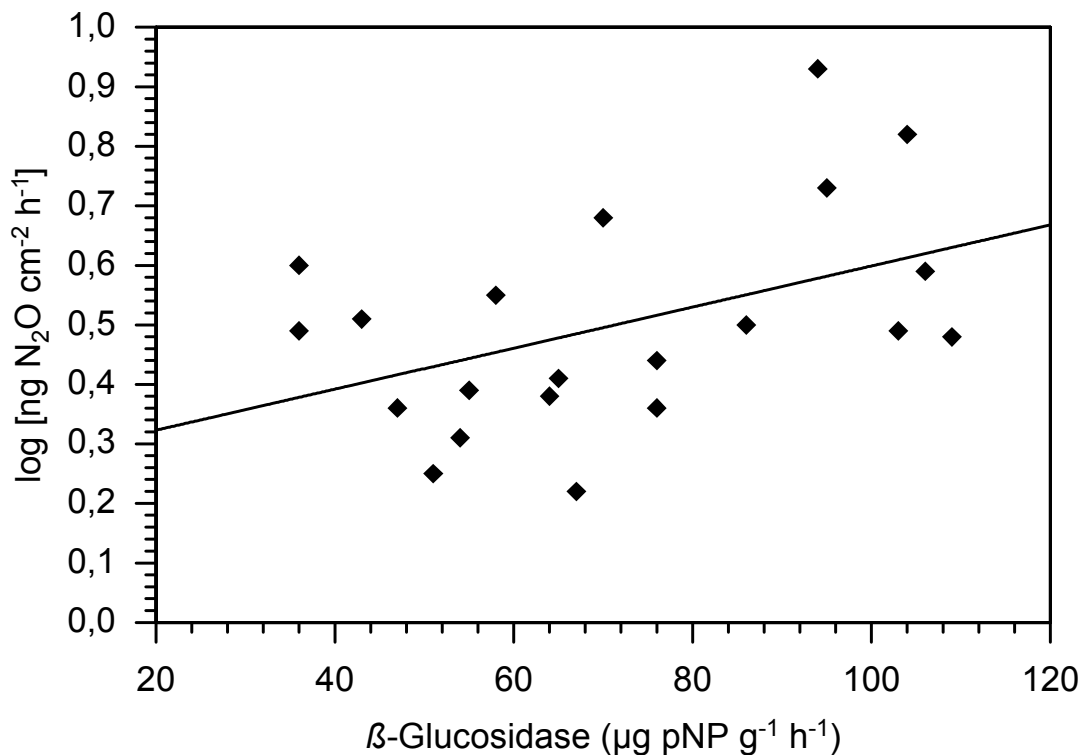


Fig. 8. (f) Linear regressions of dry season log transformed N_2O fluxes with β -Glucosidase activity [$\log \text{N}_2\text{O} = 0.003 \cdot \beta\text{-glucosidase} + 0.258$, $R^2 = 0.20$, $p = 0.042$, $n = 21$] during 2000 from a forest-to-pasture chronosequence near Santarém, Pará, Brazil.